

REVIEW ARTICLE

Dan L. Longo, M.D., *Editor*

Fibrosis — A Common Pathway to Organ Injury and Failure

Don C. Rockey, M.D., P. Darwin Bell, Ph.D., and Joseph A. Hill, M.D., Ph.D.

From the Department of Internal Medicine, Medical University of South Carolina (D.C.R., P.D.B.), and the Ralph H. Johnson Veterans Affairs Medical Center (P.D.B.) — both in Charleston; and the Departments of Internal Medicine and Molecular Biology, University of Texas Southwestern Medical Center, Dallas (J.A.H.). Address reprint requests to Dr. Rockey at the Department of Internal Medicine, Medical University of South Carolina, 96 Jonathan Lucas St., Suite 803, MSC 623, Charleston, SC 29425, or at rockey@musc.edu.

N Engl J Med 2015;372:1138-49.

DOI: 10.1056/NEJMra1300575

Copyright © 2015 Massachusetts Medical Society.

DISEASE-RELATED INJURY IN ANY ORGAN TRIGGERS A COMPLEX CASCADE of cellular and molecular responses that culminates in tissue fibrosis. Although this fibrogenic response may have adaptive features in the short term, when it progresses over a prolonged period of time, parenchymal scarring and ultimately cellular dysfunction and organ failure ensue (Fig. 1).

We and others have proposed four major phases of the fibrogenic response (Fig. 2). First is initiation of the response, driven by primary injury to the organ. The second phase is the activation of effector cells, and the third phase is the elaboration of extracellular matrix, both of which overlap with the fourth phase, during which the dynamic deposition (and insufficient resorption) of extracellular matrix promotes progression to fibrosis and ultimately to end-organ failure.

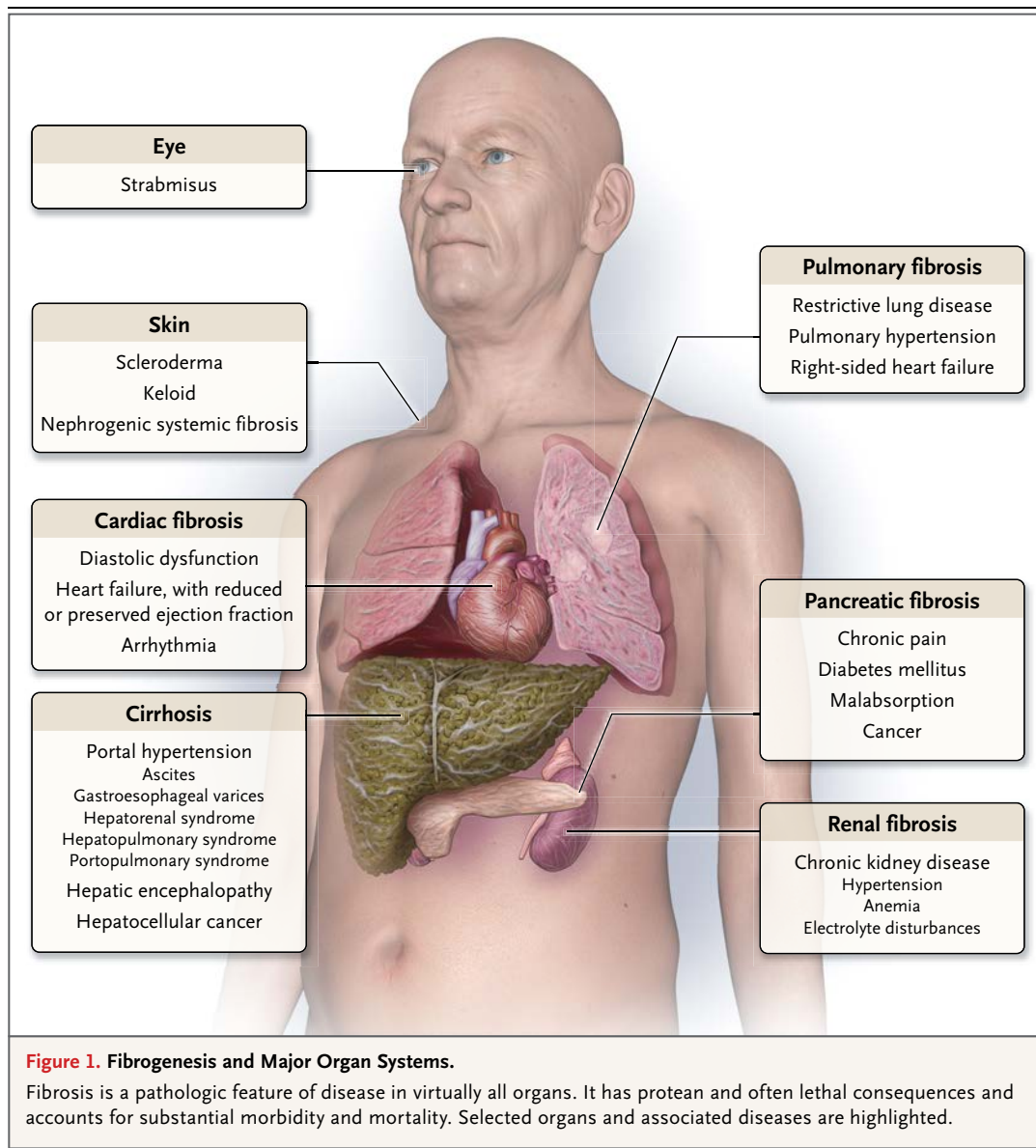
The fact that diverse diseases in different organ systems are associated with fibrotic changes suggests common pathogenic pathways (Fig. 2). This “wounding response” is orchestrated by complex activities within different cells in which specific molecular pathways have emerged. Cellular constituents include inflammatory cells (e.g., macrophages and T cells), epithelial cells, fibrogenic effector cells, endothelial cells, and others. Many different effector cells, including fibroblasts, myofibroblasts, cells derived from bone marrow, fibrocytes, and possibly cells derived from epithelial tissues (epithelial-to-mesenchymal transition) have been identified; there is some controversy regarding the identity of specific effectors in different organs. Beyond the multiple cells essential in the wounding response, core molecular pathways are critical; for example, the transforming growth factor beta (TGF- β) pathway is important in virtually all types of fibrosis.

As fibrosis progresses, myofibroblasts proliferate and sense physical and biochemical stimuli in the local environment by means of integrins and cell-surface molecules; contractile mediators trigger pathological tissue contraction. This chain of events, in turn, causes physical organ deformation, which impairs organ function. Thus, the biology of fibrogenesis is dynamic, although the degree of plasticity appears to vary from organ to organ.

Although we understand many of the cellular and molecular processes underlying fibrosis, there are few effective therapies and fewer that target fibrogenesis specifically. These facts highlight the need for a deeper comprehension of the pathogenesis of fibrogenesis and the translation of this knowledge to novel treatments.

CELLULAR AND MOLECULAR THEMES IN PATHOGENESIS

Acute and chronic inflammation often trigger fibrosis (Fig. 2). Inflammation leads to injury of resident epithelial cells and often endothelial cells, resulting in enhanced release of inflammatory mediators, including cytokines, chemokines, and others. This process leads to the recruitment of a wide range of inflammatory cells, in-



cluding lymphocytes, polymorphonuclear leukocytes, eosinophils, basophils, mast cells, and macrophages. These inflammatory cells elicit the activation of effector cells,¹ which drive the fibrogenic process. One example in which an inflammatory lesion drives this cycle is the interstitial nephritis induced by nonsteroidal antiinflammatory drugs (NSAIDs), which culminates in chronic inflammation and the activation of a fibrogenic cascade. In addition, macrophages can play a prominent role in interstitial fibrosis, often driven by the TGF- β pathway.² However, some inflammatory cells may be protective. For example,

certain populations of macrophages phagocytose apoptotic cells that promote the fibrogenic process and activate matrix-degrading metalloproteases.³

Fibroblasts and myofibroblasts have been identified as key fibrosis effectors in many organs, and as such are responsible for the synthesis of extracellular matrix proteins⁴ (Fig. 2). Controversy exists regarding the origin of these cells; for example, myofibroblasts may be derived from fibroblasts or from other mesenchymal cells, such as pericytes.⁵ Epithelial-to-mesenchymal transition, in which epithelial cells give rise to mesenchymal fibrogenic cells, may play a role. However,

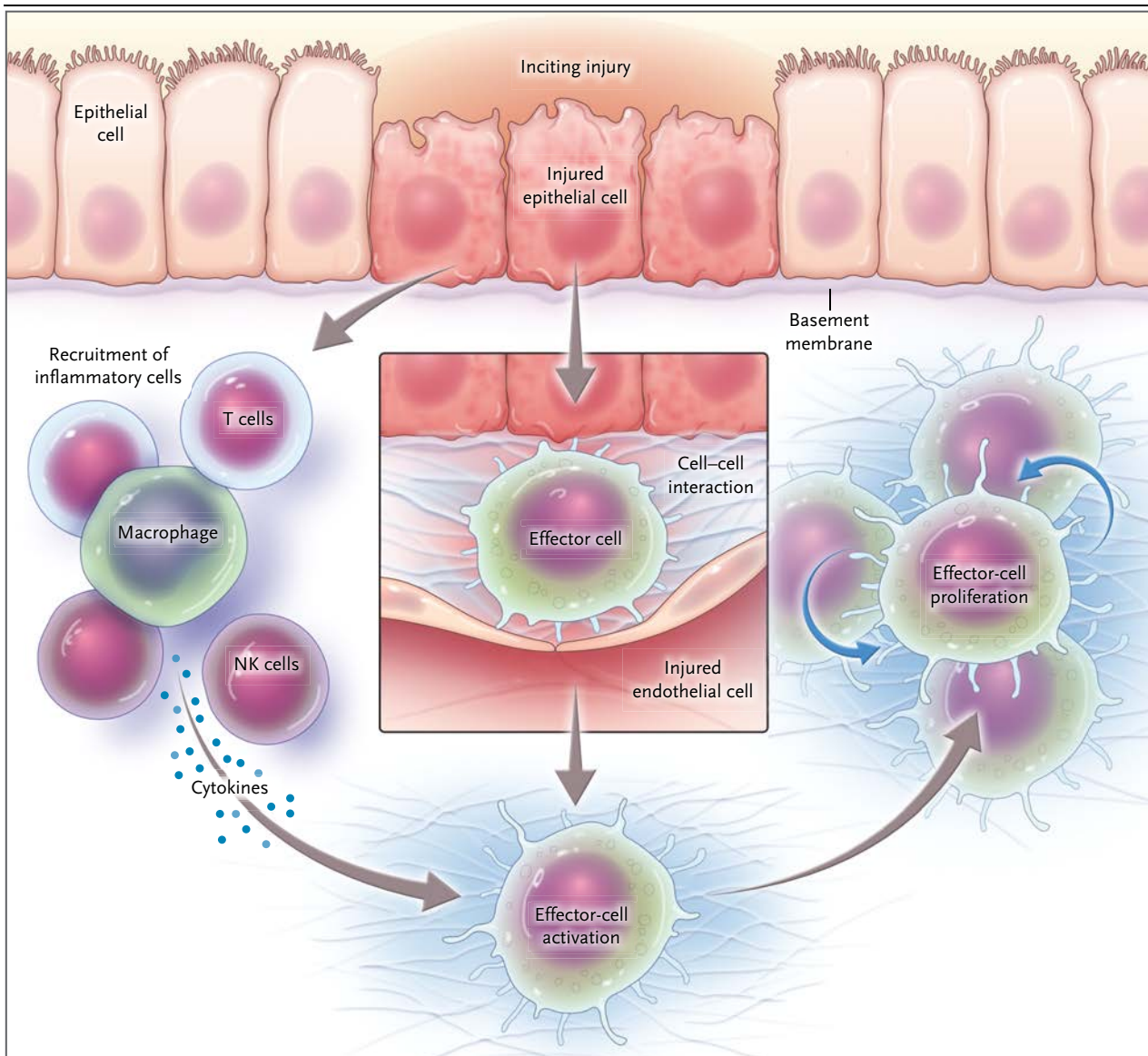


Figure 2. Cellular Injury and Fibrogenesis.

In parenchymal organs, many different types of stimuli lead to epithelial-cell injury (top), which is typically followed by an inflammatory response (shown at left). This process stimulates a fibrogenic wound-healing response that involves multiple cellular and molecular systems. At the cellular level, the recruitment of inflammatory cells is central. Inflammatory cells produce a variety of mediators, cytokines, and other factors that are responsible for the stimulation and recruitment of other cells. Key among these cells are fibrogenic effector cells; these cells are of mesenchymal origin and include fibroblasts, fibrocytes, tissue-specific pericytes and myofibroblasts, and fibroblasts derived through epithelial-to-mesenchymal transition. These effectors produce a variety of extracellular matrix proteins, which themselves may modify the wound milieu, often stimulating fibrogenic effector cells in an autocrine fashion. Indeed, in most organ systems, autocrine loops in fibrogenic effector cells are prominent. Cell-cell interactions lead to further activation of effector cells. Effector cells produce a variety of extracellular matrix proteins, peptides, cytokines, and growth factors, all of which may lead to autocrine stimulation (see the right side of the figure), typical of most organ systems. Many forms of injury also lead to the activation and transformation of other cells, such as specialized endothelial or tissue-specific cells. Injury to these cells in turn leads to a variety of downstream effects, including activation of fibrogenic effector cells. NK denotes natural killer.

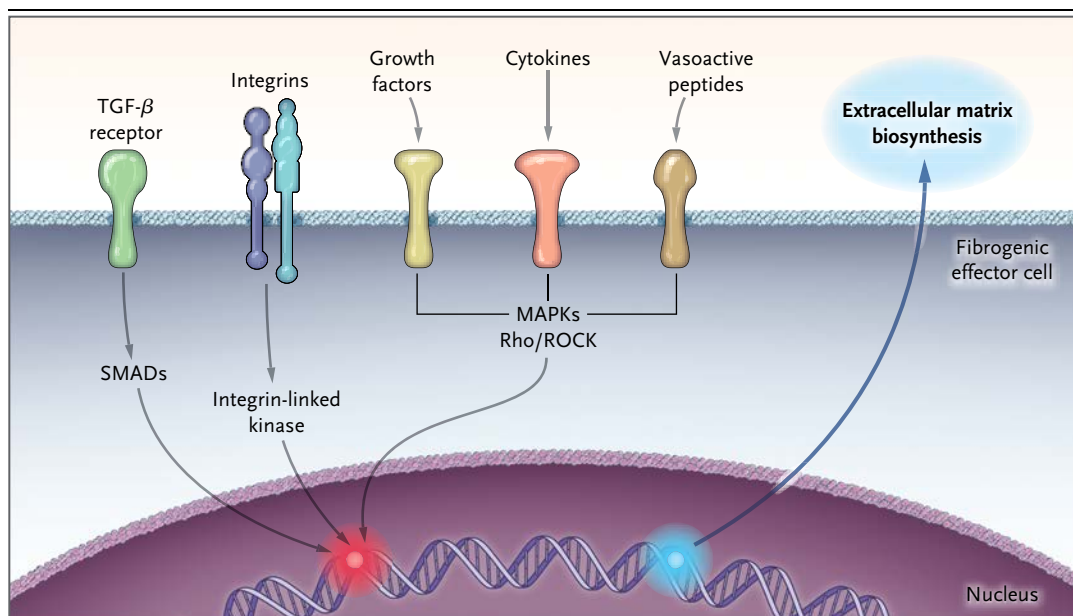


Figure 3. Molecular Pathways in Fibrosis.

Fibrogenesis in multiple tissues is effected by a number of signaling cascades. These cascades (not shown) are often triggered by the exposure of effector cells to circulating or locally produced molecules that stimulate the biosynthesis and secretion of extracellular matrix proteins. The extracellular matrix itself may also stimulate fibrogenesis through activation of integrin signaling. Examples of major pathways are shown. MAPK denotes mitogen-activated protein kinase, ROCK Rho-associated protein kinase, and TGF- β transforming growth factor β .

the importance of epithelial-to-mesenchymal transition has been actively debated.⁶⁻⁹

The matrix proteins that compose the fibrotic scar, which are highly conserved across tissues, consist predominantly of interstitial collagens (types I and III), cellular fibronectin, basement-membrane proteins such as laminin, and other, less abundant elements. In addition, myofibroblasts, which by definition are cells that express smooth-muscle proteins, including actin (ACTA2), are contractile.¹⁰ The contraction of these cells contributes to the distortion of parenchymal architecture, which promotes disease pathogenesis and tissue failure.

The molecular processes driving fibrosis are wide-ranging and complex (Fig. 3). The TGF- β cascade, which plays a major role in fibrosis, involves the binding of a ligand to a serine-threonine kinase type II receptor that recruits and phosphorylates a type I receptor. This type I receptor subsequently phosphorylates SMADs, which function as downstream effectors, typically by modulating target gene expression. Although the TGF- β superfamily, which involves multiple signaling cascades, is too complex to review in detail

here (see Massagué¹¹), its diversity highlights the complexity of the regulation of fibrosis.

TGF- β is a potent stimulator of the synthesis of extracellular matrix proteins in most fibrogenic cells. TGF- β is synthesized and secreted by inflammatory cells and by effector cells, thereby functioning in both an autocrine and paracrine fashion. The complexity of the TGF- β system is illustrated by its interactions with other cell-signaling pathways.¹² For example, TGF- β stimulates sonic hedgehog signaling in lung fibroblasts, and sonic hedgehog signaling, in turn, regulates fibroblast function.¹³ Another example of the complexity of the TGF- β system emerges from study of the extracellular activation of TGF- β in an inactive complex with a latency-associated peptide, which is subsequently activated by the $\alpha v\beta 6$ integrin.¹⁴

The molecular systems involved in fibrosis are so expansive as to preclude a detailed discussion (see Fig. 3 for an overview of several important molecular pathways). Platelet-derived growth factor (PDGF), connective-tissue growth factor (CTGF), and vasoactive peptide systems (especially angiotensin II and endothelin-1)¹⁵ play important

roles. Among vasoactive systems, endothelin plays a role in fibrosis in virtually all organ systems, acting through G-protein–coupled endothelin-A or endothelin-B cell-surface receptors or both.¹⁶ Furthermore, angiogenic pathways may be important in fibrosis.¹⁷ Finally, it is clear that integrins, which link extracellular matrix to cells, are critical in the pathogenesis of fibrosis.^{18,19}

When injury and inflammatory responses are abrogated, resorption of extracellular matrix proteins occurs, promoting organ repair. When chronic injury persists, the unremitting activation of effector cells results in the continuous deposition of extracellular matrix, progressive scarring, and organ damage. Thus, fibrogenesis involves the interplay between factors that promote the biosynthesis, deposition, and degradation of extracellular matrix proteins. Typically, matrix synthesis is counterbalanced by matrix-degrading metalloproteases.^{3,20,21} In addition, fibrogenesis is governed by pathways that eliminate effectors (e.g., by means of senescence, apoptosis, or autophagy). For example, the apoptosis of hepatic stellate cells is associated with the reversal of fibrosis.²²

An understanding of the role of genetics in the pathogenesis of fibrosis is emerging. For instance, in the kidney, fibrosis is a prominent feature of karyomegalic interstitial nephritis, which is caused by mutations in the gene encoding Fanconi anemia-associated nuclease 1 (*FANL*).²³ In the liver, *PNLAP3* is important in fibrosis that is mediated by ethanol and associated with fatty liver disease.^{24,25} A number of candidate genes may be important in cases of fibrosis that are mediated by infection with hepatitis C virus (HCV).²⁶ Mutations in *TERT* (c:2768C→T), the gene encoding telomerase reverse transcriptase, and *MUC5B*, which encodes mucin, are associated with pulmonary fibrosis.²⁷⁻²⁹

The epigenetic regulation of gene expression, which includes but is not limited to DNA methylation, post-translational modifications of the histone protein constituents of chromatin, and regulatory noncoding RNAs (e.g., microRNAs [miRs]), is important in fibrosis. For example, fibroblasts from patients with idiopathic pulmonary fibrosis have been found to have global changes in DNA methylation, changes that are not seen in fibroblasts from normal lungs.³⁰ The miRs, which play an ever-expanding role in gene regulation, are also involved in the pathogenesis of fibrosis.

In diabetic nephropathy, TGF- β promotes the expression of miR-192, which results in collagen deposition,³¹ and miR-19b regulates TGF- β signaling in hepatic stellate cells.³² In the heart, miR-21, miR-29, miR-30, and miR-133 participate in the remodeling of the myocardial matrix.³³

MECHANISMS AND ADVERSE CLINICAL EFFECTS

CARDIAC FIBROSIS

The heart undergoes extensive structural and functional remodeling in response to injury, central to which is the hypertrophy of cardiac myocytes, with excessive deposition of extracellular matrix.³⁴ Myocardial fibrosis is commonly categorized as one of two types: reactive fibrosis or replacement fibrosis. Reactive fibrosis occurs in perivascular spaces and corresponds to similar fibrogenic responses in other tissues; replacement fibrosis occurs at the site of myocyte loss.

In the heart, fibrosis is attributed to cardiac fibroblasts, the most abundant cell type in the myocardium. These cells are derived from fibroblasts that are native to the myocardium, from circulating fibroblasts, and from fibroblasts that emerge from epithelial-to-mesenchymal transition.^{35,36} All these cell types proliferate and differentiate into myofibroblasts in response to injury (Fig. 2), a process that is driven by classic factors such as TGF- β 1, endothelin-1, and angiotensin II.³⁷ Cross-talk and feedback also occur between cells — in this case, between activated fibroblasts and cardiomyocytes — which further fuel fibrogenesis.³⁸

Cardiac fibrosis contributes to both systolic and diastolic dysfunction and to perturbations of electrical excitation; it also disrupts repolarization (Fig. 1).³⁹ Proarrhythmic effects are the most prominent. Collagenous septa in the failing heart contribute to arrhythmogenesis by inducing a discontinuous slowing of conduction.⁴⁰ Areas of arrhythmogenic fibrosis slow conduction through junctions in the heterocellular gap that couple fibroblasts and cardiomyocytes.⁴¹ Endocardial breakthrough of microreentrant circuits occurs as a result of the heterogeneous spatial distribution of fibrosis⁴² and the triggering of activity caused by the depolarization of myocytes by electrically coupled myofibroblasts.⁴³

Fibrotic scarring in the heart correlates strongly with an increased incidence of arrhythmias

and sudden cardiac death.⁴⁴ For example, a 3% increase in the extracellular volume fraction of fibrous tissue (measured by means of magnetic resonance imaging after the administration of gadolinium) is associated with a 50% increase in the risk of adverse cardiac events.⁴⁵

HEPATIC FIBROSIS

Hepatic fibrosis typically results from an inflammatory process that affects hepatocytes or biliary cells. Inflammation leads to the activation of effector cells, which results in the deposition of extracellular matrix. Although a variety of effectors synthesize extracellular matrix in the liver, hepatic stellate cells appear to be the primary source of extracellular matrix. Abundant evidence suggests that the stellate cell is pericyte-like, undergoing a transformation into a myofibroblast in response to injury.¹⁰

In the liver, multiple cell types, including stellate cells, endothelial cells, Kupffer cells, bile-duct cells, and immune cells, orchestrate the cellular and molecular response to injury.⁴⁶ Numerous molecular pathways, similar to those found in other organs, are involved. A pathway that appears to be unique to the liver involves toll-like receptor 4 (TLR4)⁴⁷; TLR4 is activated on the surface of stellate cells by intestinal bacterial lipopolysaccharides derived from the gut (i.e., translocated bacteria), triggering cell activation and fibrogenesis and thereby linking fibrosis to the microbiome.⁴⁸ TLR4 expression is associated with portal inflammation and fibrosis in patients with fatty liver disease.⁴⁹

The end result of hepatic fibrogenesis is cirrhosis, an ominous parenchymal lesion that underlies a wide range of devastating complications that have adverse effects on survival (Fig. 1). Portal hypertension, a devastating result of injury, develops during the fibrogenic response after disruption of the normal interaction between sinusoidal endothelial cells and hepatic stellate cells; the resulting activation and contraction of pericyte-like stellate cells leads to sinusoidal constriction and increased intrahepatic resistance. This increase in resistance in turn activates abnormal signaling by smooth-muscle cells in mesenteric vessels. An increase in angiogenesis and collateral blood flow follows, resulting in an increase in mesenteric blood flow and a worsening of portal hypertension.⁵⁰ The major clinical sequelae of portal hypertension, variceal hemorrhage and ascites,

emerge relatively late, after the portal pressure rises to a hepatic venous pressure gradient of more than 12 mm Hg.⁵⁰

RENAL FIBROSIS

Events that initiate renal fibrosis are diverse, ranging from primary renal injury to systemic diseases.^{51,52} The kidneys are susceptible to hypertension and diabetes, the two leading causes of renal fibrosis. As is true in other organs, fibrosis of the kidney is mediated by cellular elements (e.g., inflammatory cells) and molecular elements (e.g., cytokines, TGF- β 1, CTGF, PDGF, and endothelin-1) (Fig. 2 and 3).⁵¹⁻⁵⁴ The intrarenal renin-angiotensin-aldosterone axis is particularly important in hypertension-induced fibrosis.⁵⁴

The kidney has a unique cellular architecture that consists of the glomeruli, tubules, interstitium, and capillaries. Injury at any of these sites triggers the deposition of extracellular matrix.³⁷ The location of the initial injury is an important determinant of the clinical consequences. Injuries that initially target glomeruli elicit patterns of disease that are different from those that are elicited by injuries to the tubular-Interstitial environment. For example, NSAIDs, urinary obstruction, polycystic kidney disease, and infections can provoke tubulointerstitial fibrosis,⁴³ whereas glomerular immune deposition (e.g., the deposition of IgA) leads to glomerulonephritis.⁴⁴ Glomeruli and podocytes are sensitive to systemic and local immunologic insults⁴⁵; high glomerular capillary pressure, exacerbated by systemic hypertension and diabetes, leads to proteinuria, the activation of cytokines and complement, and the infiltration of immune cells, resulting in epithelial cell and interstitial fibrosis.⁴⁶

Glomerular fibrosis, regardless of the cause, diminishes renal blood flow, which leads to hypoxia and the activation of hypoxia-inducible factor 1, which in turn triggers nephron collapse and fibrotic replacement by means of rarefaction.⁴⁷ The renal interstitium and capillaries contribute substantially to tubulointerstitial disease, as peritubular pericytes migrate into the interstitium, where they are transformed into myofibroblasts.⁴⁸

Regardless of the initiating insult, renal fibrosis leads to loss of function and organ failure (Fig. 1). Homeostasis can be maintained with a glomerular filtration rate as low as approximately 10% of the normal rate. As the mechanisms maintaining homeostasis are progressively disrupted,

anemia develops and the regulation of electrolyte balance and pH is disrupted.

PULMONARY FIBROSIS

Pulmonary fibrosis occurs in association with a wide range of diseases, including scleroderma (systemic sclerosis), sarcoidosis, and infection, and as a result of environmental exposures (e.g., silica dust or asbestos), but in most patients it is idiopathic and progressive. Idiopathic pulmonary fibrosis is characterized by progressive fibrosis without substantial inflammation.⁵⁵ Its pathogenesis appears to be unlike that of other fibrosing diseases and is poorly understood. Injury to alveolar epithelial cells activates pulmonary fibroblasts, provoking their transformation to matrix-producing myofibroblasts.⁵⁶ Activated lung fibroblasts may cause apoptosis of alveolar cells, which leads to further fibroblast activation and a vicious cycle of injury and effector-cell activation (Fig. 2). Research has focused on TGF- β signaling (Fig. 3)⁵⁷ and the interstitial pericytes present in lung fibrosis.⁵⁸

Pulmonary fibrosis is characterized by parenchymal honeycombing, reduced lung compliance, and restrictive lung function (Fig. 1). Fibrosis of the interstitial spaces hinders gas exchange, culminating in abnormal oxygenation and clinical dyspnea. Progressive pulmonary fibrosis also leads to pulmonary hypertension, right-sided heart failure, and ultimately respiratory failure.

OTHER FORMS OF FIBROSIS

Fibrosis also occurs in the joints, bone marrow, brain, eyes, intestines, peritoneum and retroperitoneum, pancreas, and skin, and in these cases is driven by typical cellular and molecular processes (Fig. 2 and 3). Retroperitoneal fibrosis is a rare condition characterized by inflammation and fibrosis in the retroperitoneal space; most cases are idiopathic, but secondary causes include drugs, infections, autoimmune and inflammatory stimuli, and radiation. Patients may present with pain, and the major clinical sequelae of this condition are related to its involvement with structures in the retroperitoneum, including arteries (leading to acute and chronic renal failure) and ureters (leading to hydronephrosis). Currently, treatment of this primary fibrosing disorder is not available. In certain cancers, fibrosis is linked to TGF- β -integrin signaling.⁵⁹ In scleroderma, the prototypical fibrosing skin disease, skin fibro-

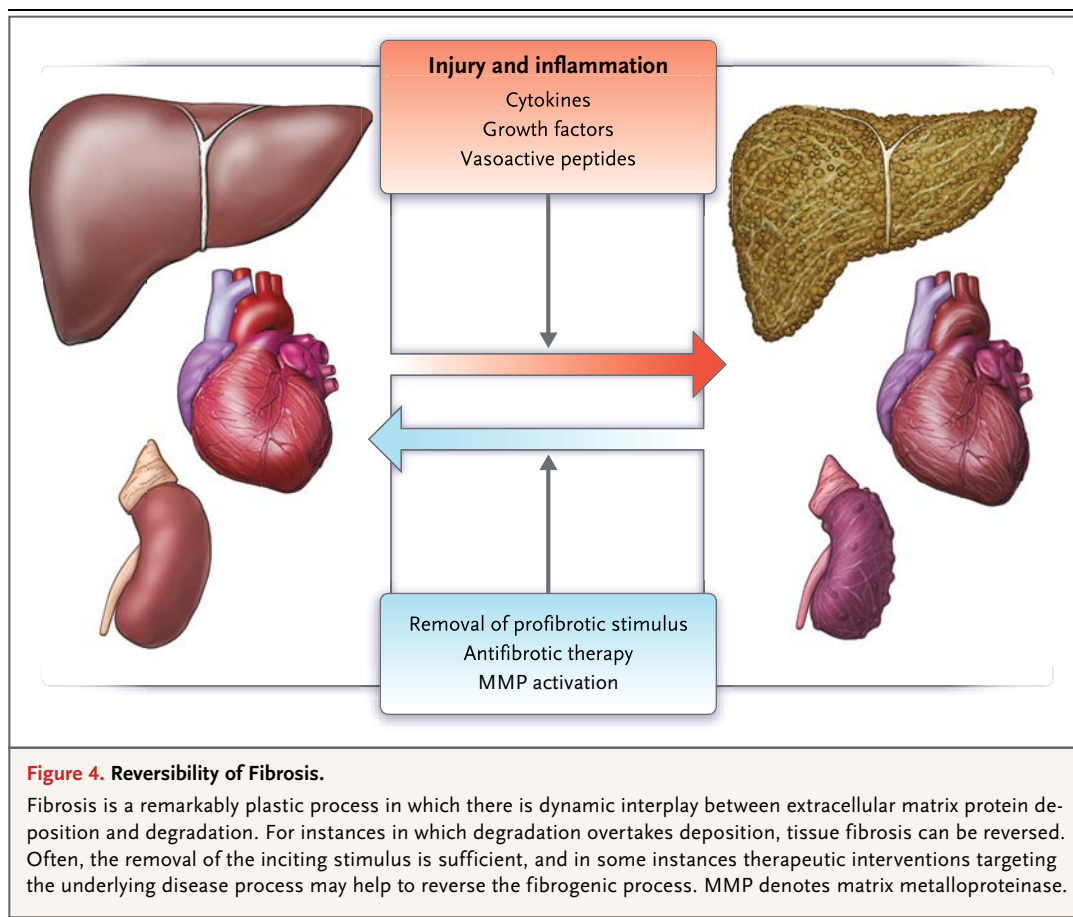
blasts and myofibroblasts are activated through the TGF- β -SMAD signaling pathway.⁶⁰ Nephrogenic systemic fibrosis, a debilitating condition that is marked by widespread organ fibrosis, occurs in patients with renal insufficiency who have been exposed to gadolinium-based contrast material. Initial systemic inflammatory-response reactions and the reaction of gadolinium (Gd³⁺) ions with circulating proteins and heavy metals lead to the deposition of insoluble elements in tissue.⁶¹ Since no effective therapies have been identified, prevention is key.⁶¹ A recently recognized IgG4-related disease appears to involve autoimmune-driven inflammation that provokes fibrosis in multiple organs, including the pancreas, retroperitoneum, lung, kidney, liver, and aorta.⁶²

THERAPY

Fibrosis and resultant organ failure account for at least one third of deaths worldwide.⁶³ Since fibrosis is common and has adverse effects in all organs, it is an attractive therapeutic target. Contrary to the widely held perception that scar tissue is permanent, the available evidence points to the highly plastic nature of organ fibrosis; it is not irreversible “scar” but an actively remodeled tissue component that can, under certain circumstances, regress. Fibrosis occurs by means of a dynamic process that involves the synthesis and deposition of extracellular matrix, and its reversal occurs by means of the elimination of effector cells and shifts in the balance of matrix synthesis and degradation (Fig. 4). Although it is not clear what pathogenic or clinical factors promote reversibility, the regression of fibrosis has been shown to lead to improved clinical outcomes. Elimination of the inciting stimulus is the first and most efficacious approach.

Fibrosis of parenchymal tissue usually progresses slowly, which suggests that therapy may be required for extended periods; slowing the progression of fibrosis may be a more realistic therapeutic goal than eliminating it. One of the challenges in assessing the therapeutic response is that there are few noninvasive means of measuring fibrosis (e.g., blood or imaging tests; biopsy is typically the most reliable approach) or monitoring the effects of therapeutic intervention.

The best indication that fibrosis is reversible and that this reversibility has positive effects on clinical outcomes is based on the treatment of



liver disease⁶⁴; in patients with cirrhosis who are infected with hepatitis B virus (HBV), antiviral therapy reduces fibrosis, reverses cirrhosis,⁶⁵ and reduces the incidence of clinical complications.⁶⁶ Pirfenidone has been shown to slow the reduction in forced vital capacity and reduce mortality, raising the possibility of a reversal in fibrosis.

Often, promising preclinical studies are not borne out in clinical trials in terms of both expected efficacy and unexpected side effects. Thus, at the current time, specific antifibrotic therapies are limited (see Table 1, which summarizes core concepts from large, completed clinical trials targeting fibrosis in humans [including both positive and negative results], and Table S1 in the Supplementary Appendix, which includes more detailed information about completed clinical trials and is available with the full text of this article at NEJM.org). Several agents targeting specific fibrotic pathways in the liver and the lung have been examined, placing the studies of the liver and lung ahead of studies of other organ systems

in terms of the goal to specifically confront fibrosis. Novel approaches to the treatment of fibrosis that are based on an extensive body of preclinical data are anticipated in the coming years (see Table S2, which highlights early phase trials that target less well established, although potentially important, pathways in fibrosis).

HEART

Pharmacologic therapies in clinical use for heart failure that target the primary underlying disease appear to have a secondary effect on fibrosis. Examples include angiotensin-converting-enzyme (ACE) inhibitors, statins, aldosterone antagonists, and emerging therapies, such as histone deacetylase inhibitors. Such therapies, which are known to promote beneficial “reverse remodeling,” ameliorate fibrosis, reduce the burden of ventricular arrhythmia, slow the rate of ventricular tachycardias,^{93,94} and reduce the incidence of sudden death.³⁹

A promising idea for the treatment of cardiac

Table 1. Pathways and Processes in Fibrogenesis and Current Treatments.*

Organ	Pathways and Processes	Diseases	Drugs	Summary of Effectiveness	Source of Data†
Heart	Aldosterone antagonism, TGF- β antagonism, RAS inhibition, cGMP inhibition, inhibition of cholesterol synthesis, inhibition of Na-K-Cl cotransporter	Heart failure, cardiomyopathy, hypertrophic cardiomyopathy, cardiomyopathy induced by type 2 diabetes, heart failure or cardiomyopathy induced by hypertension	Spironolactone, eplerenone, canrenone, pirfenidone, sildenafil, statins, ACE inhibitors, ARBs, torsemide, MRAs	ACE inhibitors, ARBs, and MRAs are associated with decreased fibrosis on MRI and decreased arrhythmogenesis (the latter suggests effects of drugs on fibrosis)	Kosmala et al., ⁶⁷ Giannetta et al., ⁶⁸ Antonopoulos et al., ⁶⁹ Roubille et al., ⁷⁰ TORAFIC Investigators Group ⁷¹
Liver	RAS inhibition, inhibition of collagen synthesis, inhibition of effector-cell fibrogenesis, inhibition of oxidative stress, signaling of PPAR γ -agonists	Many diseases of the liver	ACE inhibitors, ARBs, colchicine, interferon γ -1b, vitamin E, pioglitazone, farglitazar	Specific antifibrotic agents listed have generally been ineffective in halting or reversing fibrosis	Sanyal et al., ⁷² Kim et al., ⁷³ Kershenovich et al., ⁷⁴ Morgan et al., ⁷⁵ Muir et al., ⁷⁶ Pockros et al., ⁷⁷ McHutchison et al. ⁷⁸
Kidney	RAS inhibition, aldosterone antagonism, TGF- β antagonism, Nrf2 pathway	Primarily renal diseases related to hypertension or diabetes	ACE inhibitors, ARBs, spironolactone, pirfenidone, bardoxolone	ACE inhibitors and ARBs are moderately effective in slowing progression of diabetic nephropathy (indirectly suggesting effects on fibrosis)	Lambers Heerspink et al., ⁷⁹ Ruggerenti et al., ⁸⁰ Bonventre, ⁸¹ Guney et al., ⁸² Sharma et al., ⁸³ de Zeeuw et al. ⁸⁴
Lung	TGF- β antagonism, direct inhibition of effector-cell fibrogenesis, multikinase inhibition, inhibition of oxidative stress	Primarily idiopathic pulmonary fibrosis	Pirfenidone, interferon γ -1b, bosentan, ambrisentan, macitentan, nintedanib, acetylcysteine	Pirfenidone and nintedanib led to improvements in clinical outcomes	Raghu et al., ⁸⁵⁻⁸⁷ King et al., ⁸⁸ Richeldi et al., ⁸⁹ Martinez et al. ⁹⁰
Skin	Endothelin-receptor antagonism, multikinase inhibition	Scleroderma, nephrogenic systemic fibrosis	Bosentan, imatinib mesylate	Small studies show modest effects	Kuhn et al., ⁹¹ Kay and High ⁹²

* ACE denotes angiotensin-converting enzyme, ARB angiotensin-receptor blocker, cGMP cyclic guanosine monophosphate, MRA mineralocorticoid-receptor antagonist, MRI magnetic resonance imaging, Nrf2 nuclear factor erythroid 2–related factor, PPAR peroxisome proliferator-activated receptor, RAS renin–angiotensin system, and TGF- β transforming growth factor beta.

† Detailed information about specific trials is provided in Table S1 in the Supplementary Appendix.

fibrosis is based on the premise that cardiac fibroblasts can be reprogrammed into cardiomyocyte-like cells^{95,96} (Table S2 in the Supplementary Appendix) that would promote normal tissue regeneration; in a murine model of myocardial infarction and fibrosis, the retroviral expression of specific transcription factors in the myocardium reprogrammed cells that acquired spontaneous contractile and electrophysiological properties resembling those of cardiomyocytes, leading to global improvements in contractile function. It is not yet known whether this type of therapy can be used in humans.

KIDNEY

Like the therapies used to treat cardiac fibrosis, those typically used to prevent renal fibrosis target the underlying disease processes and as such involve the treatment of hypertension and diabetes. One target is the renin–angiotensin system.

This approach involves the use of ACE inhibitors and angiotensin-receptor blockers that ameliorate renal damage and fibrosis through multiple pathways, including the suppression of the actions of TGF- β .⁷⁹ Therapies based on the antagonism of aldosterone that make use of mineralocorticoid receptor antagonists have been shown to inhibit or slow the progression of fibrosis in humans.⁹⁷ Novel approaches to the treatment of fibrosis of the kidneys include those that target bone morphogenetic protein-7, NADPH oxidase (NOX) (NOX1 and NOX4), and the SMAD3 and SMAD4 pathways (Table S2 in the Supplementary Appendix).⁹⁸

LIVER

The process of hepatic fibrosis is dynamic. Since hepatocytes are capable of regeneration, liver fibrosis may be especially amenable to therapeutic intervention, and even cirrhosis can be re-

versed.^{66,99-101} Eradication of HCV infection, antiviral therapy for HBV infection, glucocorticoid therapy for autoimmune hepatitis, phlebotomy for hemochromatosis, relief of biliary obstruction, and cessation of alcohol consumption in alcoholic hepatitis each clearly reverses fibrotic change, and many of these treatments improve clinical outcomes.^{66,99,100,102}

A number of potential antifibrotic therapies targeting specific pathways have been studied in human liver disease (Table S2 in the Supplementary Appendix). For example, colchicine suppresses collagen secretion and theoretically prevents fibrosis.⁴⁶ Interferon γ -1b and the peroxisome proliferator-activated receptor γ ligand, farglitar, which inhibit stellate cell-mediated fibrogenesis, were studied in patients infected with HCV that was unresponsive to primary antiviral therapy, but no beneficial effects on fibrosis were noted.⁴⁶ Other agents, including polyene-phosphatidyl choline, silymarin, and ursodeoxycholic acid, have similarly shown no benefit.⁴⁶ Vitamin E had modest effects on histologic fibrosis in patients with nonalcoholic steatohepatitis.⁷²

LUNG

The lung presents special challenges with regard to therapy targeting fibrosis. On the one hand, the lung has easily measured clinical features that allow for assessment of lung function, a surrogate for fibrosis. On the other hand, pulmonary fibrosis appears to be less dynamic than fibrosis occurring in other organ systems. Multiple therapies have been tested for pulmonary fibrosis, especially for idiopathic pulmonary fibrosis. Interferon γ -1b showed efficacy in preclinical studies but showed no benefit in human clinical trials.⁸⁵ Endothelin-receptor antagonists have also

showed no benefit. Pirfenidone, a pyridone derivative with antiinflammatory and antifibrotic effects that is available in oral form, reduced disease progression and increased survival in patients with idiopathic pulmonary fibrosis. Its mechanism of action is not completely understood, but it presumably has effects on TGF- β production.⁸⁸ Nintedanib, a multikinase inhibitor, slowed disease progression in a similar cohort.⁸⁹ It is important to note that the outcomes in these trials were based on clinical results; reductions in lung fibrosis per se have not been definitively demonstrated.

FUTURE DIRECTIONS

Fibrosis is a hallmark of pathologic remodeling in numerous tissues and a contributor to clinical disease. There is a great deal of interest in identifying means of slowing, arresting, or even reversing the progression of tissue fibrogenesis. Thus, it is important to understand the central mechanisms underlying the fibrogenic process. Common themes implicating conserved cellular and molecular pathways have emerged. A major conserved cellular element is the activated fibroblast, also known as a myofibroblast, which produces abundant amounts of extracellular matrix. Some of the major conserved molecular processes involve TGF- β , PDGF, CTGF, vasoactive compounds (endothelin-1 and angiotensin II), and integrin-extracellular matrix signaling pathways. The fact that tissue fibrosis is remarkably plastic suggests that many of these major elements of disease pathogenesis may emerge as targets of novel therapeutic interventions.

Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

REFERENCES

- Wynn TA. Fibrotic disease and the T(H)1/T(H)2 paradigm. *Nat Rev Immunol* 2004;4:583-94.
- Meng XM, Nikolic-Paterson DJ, Lan HY. Inflammatory processes in renal fibrosis. *Nat Rev Nephrol* 2014;10:493-503.
- Ramachandran P, Iredale JP. Macrophages: central regulators of hepatic fibrogenesis and fibrosis resolution. *J Hepatol* 2012;56:1417-9.
- Hinz B, Phan SH, Thannickal VJ, Galli A, Bochaton-Piallat ML, Gabbiani G. The myofibroblast: one function, multiple origins. *Am J Pathol* 2007;170:1807-16.
- Kida Y, Duffield JS. Pivotal role of pericytes in kidney fibrosis. *Clin Exp Pharmacol Physiol* 2011;38:467-73.
- Carew RM, Wang B, Kantharidis P. The role of EMT in renal fibrosis. *Cell Tissue Res* 2012;347:103-16.
- Taura K, Miura K, Iwaisako K, et al. Hepatocytes do not undergo epithelial-mesenchymal transition in liver fibrosis in mice. *Hepatology* 2010;51:1027-36.
- Rock JR, Barkauskas CE, Crompton MJ, et al. Multiple stromal populations contribute to pulmonary fibrosis without evidence for epithelial to mesenchymal transition. *Proc Natl Acad Sci U S A* 2011;108(52):E1475-E1483.
- LeBleu VS, Taduri G, O'Connell J, et al. Origin and function of myofibroblasts in kidney fibrosis. *Nat Med* 2013;19:1047-53.
- Rockey DC, Housset CN, Friedman SL. Activation-dependent contractility of rat hepatic lipocytes in culture and in vivo. *J Clin Invest* 1993;92:1795-804.
- Massagué J. TGF β signalling in context. *Nat Rev Mol Cell Biol* 2012;13:616-30.
- Guo X, Wang XF. Signaling cross-talk between TGF-beta/BMP and other pathways. *Cell Res* 2009;19:71-88.
- Cigna N, Farrokhi Moshai E, Brayer S,

- et al. The hedgehog system machinery controls transforming growth factor- β -dependent myofibroblastic differentiation in humans: involvement in idiopathic pulmonary fibrosis. *Am J Pathol* 2012;181:2126-37.
14. Munger JS, Huang X, Kawakatsu H, et al. The integrin α v β 6 binds and activates latent TGF β 1: a mechanism for regulating pulmonary inflammation and fibrosis. *Cell* 1999;96:319-28.
15. Wynn TA. Common and unique mechanisms regulate fibrosis in various fibroproliferative diseases. *J Clin Invest* 2007;117:524-9.
16. Khimji AK, Rockey DC. Endothelin — biology and disease. *Cell Signal* 2010;22:1615-25.
17. Johnson A, DiPietro LA. Apoptosis and angiogenesis: an evolving mechanism for fibrosis. *FASEB J* 2013;27:3893-901.
18. Levine D, Rockey DC, Milner TA, Breuss JM, Fallon JT, Schnapp LM. Expression of the integrin α 8 β 1 during pulmonary and hepatic fibrosis. *Am J Pathol* 2000;156:1927-35.
19. Henderson NC, Arnold TD, Katamura Y, et al. Targeting of α v integrin identifies a core molecular pathway that regulates fibrosis in several organs. *Nat Med* 2013;19:1617-24.
20. Shechter R, Raposo C, London A, Sagi I, Schwartz M. The glial scar-monocyte interplay: a pivotal resolution phase in spinal cord repair. *PLoS One* 2011;6(12):e27969.
21. Du X, Shimizu A, Masuda Y, et al. Involvement of matrix metalloproteinase-2 in the development of renal interstitial fibrosis in mouse obstructive nephropathy. *Lab Invest* 2012;92:1149-60.
22. Fallowfield JA, Mizuno M, Kendall TJ, et al. Scar-associated macrophages are a major source of hepatic matrix metalloproteinase-13 and facilitate the resolution of murine hepatic fibrosis. *J Immunol* 2007;178:5288-95.
23. Zhou W, Otto EA, Cluckey A, et al. FAN1 mutations cause karyomegalic interstitial nephritis, linking chronic kidney failure to defective DNA damage repair. *Nat Genet* 2012;44:910-5.
24. Tian C, Stokowski RP, Kershenovich D, Ballinger DG, Hinds DA. Variant in PNPLA3 is associated with alcoholic liver disease. *Nat Genet* 2010;42:21-3.
25. Romeo S, Kozlitina J, Xing C, et al. Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet* 2008;40:1461-5.
26. Huang H, Shiffman ML, Friedman S, et al. A 7 gene signature identifies the risk of developing cirrhosis in patients with chronic hepatitis C. *Hepatology* 2007;46:297-306.
27. Gansner JM, Rosas IO, Ebert BL. Pulmonary fibrosis, bone marrow failure, and telomerase mutation. *N Engl J Med* 2012;366:1551-3.
28. Hunninghake GM, Hatabu H, Okajima Y, et al. MUC5B promoter polymorphism and interstitial lung abnormalities. *N Engl J Med* 2013;368:2192-200.
29. Fingerlin TE, Murphy E, Zhang W, et al. Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. *Nat Genet* 2013;45:613-20.
30. Huang SK, Scruggs AM, McEachin RC, White ES, Peters-Golden M. Lung fibroblasts from patients with idiopathic pulmonary fibrosis exhibit genome-wide differences in DNA methylation compared to fibroblasts from nonfibrotic lung. *PLoS One* 2014;9(9):e107055.
31. Putta S, Lanting L, Sun G, Lawson G, Kato M, Natarajan R. Inhibiting microRNA-192 ameliorates renal fibrosis in diabetic nephropathy. *J Am Soc Nephrol* 2012;23:458-69.
32. Lakner AM, Steuerwald NM, Walling TL, et al. Inhibitory effects of microRNA 19b in hepatic stellate cell-mediated fibrogenesis. *Hepatology* 2012;56:300-10.
33. van Rooij E, Olson EN. Searching for miRNAs in cardiac fibrosis. *Circ Res* 2009;104:138-40.
34. Hill JA, Olson EN. Cardiac plasticity. *N Engl J Med* 2008;358:1370-80.
35. Zeisberg EM, Tarnavski O, Zeisberg M, et al. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat Med* 2007;13:952-61.
36. Moore-Morris T, Guimarães-Camboa N, Banerjee I, et al. Resident fibroblast lineages mediate pressure overload-induced cardiac fibrosis. *J Clin Invest* 2014;124:2921-34.
37. Burchfield JS, Xie M, Hill JA. Pathological ventricular remodeling: mechanisms: part 1 of 2. *Circulation* 2013;128:388-400.
38. Martin ML, Blaxall BC. Cardiac intercellular communication: are myocytes and fibroblasts fair-weather friends? *J Cardiovasc Transl Res* 2012;5:768-82.
39. Spinale FG. Myocardial matrix remodeling and the matrix metalloproteinases: influence on cardiac form and function. *Physiol Rev* 2007;87:1285-342.
40. Spach MS, Boineau JP. Microfibrosis produces electrical load variations due to loss of side-to-side cell connections: a major mechanism of structural heart disease arrhythmias. *Pacing Clin Electrophysiol* 1997;20:397-413.
41. Miragoli M, Gaudesius G, Rohr S. Electrotonic modulation of cardiac impulse conduction by myofibroblasts. *Circ Res* 2006;98:801-10.
42. Tanaka K, Zlochiver S, Vikstrom KL, et al. Spatial distribution of fibrosis governs fibrillation wave dynamics in the posterior left atrium during heart failure. *Circ Res* 2007;101:839-47.
43. Miragoli M, Salvarani N, Rohr S. Myofibroblasts induce ectopic activity in cardiac tissue. *Circ Res* 2007;101:755-8.
44. Wu KC, Weiss RG, Thiemann DR, et al. Late gadolinium enhancement by cardiovascular magnetic resonance heralds an adverse prognosis in nonischemic cardiomyopathy. *J Am Coll Cardiol* 2008;51:2414-21.
45. Wong TC, Piehler K, Meier CG, et al. Association between extracellular matrix expansion quantified by cardiovascular magnetic resonance and short-term mortality. *Circulation* 2012;126:1206-16.
46. Rockey DC. Translating an understanding of the pathogenesis of hepatic fibrosis to novel therapies. *Clin Gastroenterol Hepatol* 2013;11(3):224-31.
47. Seki E, De Minicis S, Osterreicher CH, et al. TLR4 enhances TGF- β signaling and hepatic fibrosis. *Nat Med* 2007;13:1324-32.
48. Fouts DE, Torralba M, Nelson KE, Brenner DA, Schnabl B. Bacterial translocation and changes in the intestinal microbiome in mouse models of liver disease. *J Hepatol* 2012;56:1283-92.
49. Vespasiani-Gentilucci U, Carotti S, Perrone G, et al. Hepatic toll-like receptor 4 expression is associated with portal inflammation and fibrosis in patients with NAFLD. *Liver Int* 2015;35:569-81.
50. Sanyal AJ, Bosch J, Blei A, Arroyo V. Portal hypertension and its complications. *Gastroenterology* 2008;134:1715-28.
51. Liu Y. Cellular and molecular mechanisms of renal fibrosis. *Nat Rev Nephrol* 2011;7:684-96.
52. Kaissling B, Lehir M, Kriz W. Renal epithelial injury and fibrosis. *Biochim Biophys Acta* 2013;1832:931-9.
53. Chen J, Chen JK, Nagai K, et al. EGFR signaling promotes TGF β -dependent renal fibrosis. *J Am Soc Nephrol* 2012;23:215-24.
54. Mezzano SA, Ruiz-Ortega M, Egidio J. Angiotensin II and renal fibrosis. *Hypertension* 2001;38:635-8.
55. Thannickal VJ, Toews GB, White ES, Lynch JP III, Martinez FJ. Mechanisms of pulmonary fibrosis. *Annu Rev Med* 2004;55:395-417.
56. Sakai N, Tager AM. Fibrosis of two: epithelial cell-fibroblast interactions in pulmonary fibrosis. *Biochim Biophys Acta* 2013;1832:911-21.
57. Wynn TA. Integrating mechanisms of pulmonary fibrosis. *J Exp Med* 2011;208:1339-50.
58. Hung C, Linn G, Chow YH, et al. Role of lung pericytes and resident fibroblasts in the pathogenesis of pulmonary fibrosis. *Am J Respir Crit Care Med* 2013;188:820-30.
59. Margadant C, Sonnenberg A. Integrin-TGF- β crosstalk in fibrosis, cancer and wound healing. *EMBO Rep* 2010;11:97-105.
60. Jinnin M. Mechanisms of skin fibrosis in systemic sclerosis. *J Dermatol* 2010;37:11-25.

61. Swaminathan S, Horn TD, Pellowski D, et al. Nephrogenic systemic fibrosis, gadolinium, and iron mobilization. *N Engl J Med* 2007;357:720-2.
62. Umehara H, Okazaki K, Masaki Y, et al. A novel clinical entity, IgG4-related disease (IgG4RD): general concept and details. *Mod Rheumatol* 2012;22:1-14.
63. Zeisberg M, Kalluri R. Cellular mechanisms of tissue fibrosis: 1. common and organ-specific mechanisms associated with tissue fibrosis. *Am J Physiol Cell Physiol* 2013;304:C216-C225.
64. Liaw YF. Reversal of cirrhosis: an achievable goal of hepatitis B antiviral therapy. *J Hepatol* 2013;59:880-1.
65. Marcellin P, Gane E, Buti M, et al. Regression of cirrhosis during treatment with tenofovir disoproxil fumarate for chronic hepatitis B: a 5-year open-label follow-up study. *Lancet* 2013;381:468-75.
66. Liaw YF, Sung JJ, Chow WC, et al. Lamivudine for patients with chronic hepatitis B and advanced liver disease. *N Engl J Med* 2004;351:1521-31.
67. Kosmala W, Przewlocka-Kosmala M, Szczepanik-Osadnik H, Mysiak A, O'Moore-Sullivan T, Marwick TH. A randomized study of the beneficial effects of aldosterone antagonism on LV function, structure, and fibrosis markers in metabolic syndrome. *JACC Cardiovasc Imaging* 2011;4:1239-49.
68. Giannetta E, Isidori AM, Galea N, et al. Chronic inhibition of cGMP phosphodiesterase 5A improves diabetic cardiomyopathy: a randomized, controlled clinical trial using magnetic resonance imaging with myocardial tagging. *Circulation* 2012;125:2323-33.
69. Antonopoulos AS, Margaritis M, Lee R, Channon K, Antoniadou C. Statins as anti-inflammatory agents in atherosclerosis: molecular mechanisms and lessons from the recent clinical trials. *Curr Pharm Des* 2012;18:1519-30.
70. Roubille F, Busseuil D, Merlet N, Kritikou EA, Rhéaume E, Tardif JC. Investigational drugs targeting cardiac fibrosis. *Expert Rev Cardiovasc Ther* 2014;12:111-25.
71. TORAFIC Investigators Group. Effects of prolonged-release torasemide versus furosemide on myocardial fibrosis in hypertensive patients with chronic heart failure: a randomized, blinded-end point, active-controlled study. *Clin Ther* 2011;33:1204.e3-13.e3.
72. Sanyal AJ, Chalasani N, Kowdley KV, et al. Pioglitazone, vitamin E, or placebo for nonalcoholic steatohepatitis. *N Engl J Med* 2010;362:1675-85.
73. Kim MY, Cho MY, Baik SK, et al. Beneficial effects of candesartan, an angiotensin-blocking agent, on compensated alcoholic liver fibrosis — a randomized open-label controlled study. *Liver Int* 2012;32:977-87.
74. Kershenobich D, Vargas F, Garcia-Tsao G, Perez Tamayo R, Gent M, Rojkind M. Colchicine in the treatment of cirrhosis of the liver. *N Engl J Med* 1988;318:1709-13.
75. Morgan TRWD, Weiss DG, Nemchausky B, et al. Colchicine treatment of alcoholic cirrhosis: a randomized, placebo-controlled clinical trial of patient survival. *Gastroenterology* 2005;128:882-90.
76. Muir AJ, Sylvestre PB, Rockey DC. Interferon gamma-1b for the treatment of fibrosis in chronic hepatitis C infection. *J Viral Hepat* 2006;13:322-8.
77. Pockros PJ, Jeffers L, Afdhal N, et al. Final results of a double-blind, placebo-controlled trial of the antifibrotic efficacy of interferon-gamma1b in chronic hepatitis C patients with advanced fibrosis or cirrhosis. *Hepatology* 2007;45:569-78.
78. McHutchison J, Goodman Z, Patel K, et al. Farglitazar lacks antifibrotic activity in patients with chronic hepatitis C infection. *Gastroenterology* 2010;138:1365-73.
79. Lambers Heerspink HJ, de Borst MH, Bakker SJ, Navis GJ. Improving the efficacy of RAAS blockade in patients with chronic kidney disease. *Nat Rev Nephrol* 2013;9:112-21.
80. Ruggenenti P, Cravedi P, Remuzzi G. The RAAS in the pathogenesis and treatment of diabetic nephropathy. *Nat Rev Nephrol* 2010;6:319-30.
81. Bonventre JV. Can we target tubular damage to prevent renal function decline in diabetes? *Semin Nephrol* 2012;32:452-62.
82. Guney I, Selcuk NY, Altintepe L, Atalay H, Başarali MK, Büyükbay S. Antifibrotic effects of aldosterone receptor blocker (spironolactone) in patients with chronic kidney disease. *Ren Fail* 2009;31:779-84.
83. Sharma K, Ix JH, Mathew AV, et al. Pirfenidone for diabetic nephropathy. *J Am Soc Nephrol* 2011;22:1144-51.
84. de Zeeuw D, Akizawa T, Audhya P, et al. Bardoxolone methyl in type 2 diabetes and stage 4 chronic kidney disease. *N Engl J Med* 2013;369:2492-503.
85. Raghu G, Brown KK, Bradford WZ, et al. A placebo-controlled trial of interferon gamma-1b in patients with idiopathic pulmonary fibrosis. *N Engl J Med* 2004;350:125-33.
86. Raghu G, Behr J, Brown KK, et al. Treatment of idiopathic pulmonary fibrosis with ambrisentan: a parallel, randomized trial. *Ann Intern Med* 2013;158:641-9.
87. Raghu G, Million-Rousseau R, Morganti A, et al. Macitentan for the treatment of idiopathic pulmonary fibrosis: the randomised controlled MUSIC trial. *Eur Respir J* 2013;42:1622-32.
88. King TE Jr, Bradford WZ, Castro-Bernardini S, et al. A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2083-92.
89. Richeldi L, du Bois RM, Raghu G, et al. Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2071-82.
90. Martinez FJ, de Andrade JA, Anstrom KJ, et al. Randomized trial of acetylcysteine in idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2093-101.
91. Kuhn A, Haust M, Ruland V, et al. Effect of bosentan on skin fibrosis in patients with systemic sclerosis: a prospective, open-label, non-comparative trial. *Rheumatology (Oxford)* 2010;49:1336-45.
92. Kay J, High WA. Imatinib mesylate treatment of nephrogenic systemic fibrosis. *Arthritis Rheum* 2008;58:2543-8.
93. Massare J, Berry JM, Luo X, et al. Diminished cardiac fibrosis in heart failure is associated with altered ventricular arrhythmia phenotype. *J Cardiovasc Electrophysiol* 2010;21:1031-7.
94. Dimas V, Ayers C, Daniels J, Joglar JA, Hill JA, Naseem RH. Spironolactone therapy is associated with reduced ventricular tachycardia rate in patients with cardiomyopathy. *Pacing Clin Electrophysiol* 2011;34:309-14.
95. Song K, Nam YJ, Luo X, et al. Heart repair by reprogramming non-myocytes with cardiac transcription factors. *Nature* 2012;485:599-604.
96. Qian L, Huang Y, Spencer CI, et al. In vivo reprogramming of murine cardiac fibroblasts into induced cardiomyocytes. *Nature* 2012;485:593-8.
97. Bianchi S, Bigazzi R, Campese VM. Antagonists of aldosterone and proteinuria in patients with CKD: an uncontrolled pilot study. *Am J Kidney Dis* 2005;46:45-51.
98. Tampe D, Zeisberg M. Potential approaches to reverse or repair renal fibrosis. *Nat Rev Nephrol* 2014;10:226-37.
99. Dufour JF, DeLellis R, Kaplan MM. Reversibility of hepatic fibrosis in autoimmune hepatitis. *Ann Intern Med* 1997;127:981-5.
100. Hammel P, Couvelard A, O'Toole D, et al. Regression of liver fibrosis after biliary drainage in patients with chronic pancreatitis and stenosis of the common bile duct. *N Engl J Med* 2001;344:418-23.
101. Berenguer J, Alvarez-Pellicer J, Martín PM, et al. Sustained virological response to interferon plus ribavirin reduces liver-related complications and mortality in patients coinfecting with human immunodeficiency virus and hepatitis C virus. *Hepatology* 2009;50:407-13.
102. Maylin S, Martinot-Peignoux M, Moucari R, et al. Eradication of hepatitis C virus in patients successfully treated for chronic hepatitis C. *Gastroenterology* 2008;135:821-9.

Copyright © 2015 Massachusetts Medical Society.